

Note

The structure of the chemical repeating-unit of the O-specific polysaccharide chain of *Shigella boydii* 6 lipopolysaccharide

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Bacterial lipopolysaccharides of the *Shigella* genus, unlike those of the *Salmonella*¹, have not been studied in detail. The structures proposed for *Sh flexneri* lipopolysaccharides² were recently reinvestigated³. We have chemotyped the *Sh dysenteriae*⁴ and *Sh boydii*⁵ serotypes on the basis of the monosaccharide composition of the respective lipopolysaccharides. We now report data on the structure of the O-specific chain of *Sh boydii* 6 lipopolysaccharide.

Isolation of the O-specific polysaccharide by mild hydrolysis of the lipopolysaccharide⁶ and its characterization and monosaccharide composition have been described previously⁷. The polysaccharide is composed of D-galactose, D-mannose, 2-acetamido-2-deoxy-D-galactose, and D-glucuronic acid residues in the ratios 1:2:1:1. When the polysaccharide was methylated (Hakomori⁸) and then methanolysed, and the product mixture subjected to ion-exchange chromatography, the methyl glycosides of partially methylated amino and neutral sugars were obtained. The former was identified as methyl 2-deoxy-4,6-di-O-methyl-2-methylamino-D-galactoside, since the mass spectrum of its *N,O*-diacetyl derivative was closely similar to that of the corresponding *gluco* isomer, prepared from benzyl 2-acetamido-2-deoxy-3-O-β-D-galactopyranosyl-α-D-glucopyranoside⁹. Both mass spectra were in agreement with the literature data¹⁰. The methyl glycosides of the partially methylated, neutral sugars were converted into the corresponding alditol acetates by hydrolysis, followed by reduction with borohydride and acetylation. Glc and glc-ms of the products revealed derivatives of 3,4,6-tri-O-methylmannose, 2,3,4-tri-O-methylmannose, and 2,6-di-O-methylgalactose in the ratios 1:1:0.7. Methylation analysis of the carboxyl-reduced polysaccharide⁷ gave 2,3,4,6-tetra-O-methylglucitol diacetate together with the above components in approximately equivalent amounts.

That glucuronic acid was linked to galactose followed from the isolation of a glucuronosylgalactose upon acid hydrolysis of the polysaccharide⁷. Base-catalyzed

Methylation analysis of the polysaccharide — Using the conventional procedure⁸, the methylated polysaccharide (~2 mg) was heated in a sealed tube with 1 ml of M hydrogen chloride in methanol for 20 h at 100°. The solution was then concentrated to dryness and a solution of the residue in 50% aqueous methanol was passed through a column (0.7 × 5 cm) of Dowex-50(H⁺) resin by elution with 30 ml of 50% aqueous methanol to give the methyl glycosides of the partially methylated, neutral monosaccharides. The eluate was concentrated to dryness, the residue was heated with 1 ml of 85% formic acid for 1 h at 100°, the hydrolysate was concentrated, and the residue was heated with 1 ml of 0.3M hydrochloric acid for 16 h at 100°. The resulting sugars were converted into the corresponding alditol acetates by the conventional method. G l c and g l c -m s revealed three peaks, with *T* values (relative to 1,5-di-*O*-acetyl-2,3,4,6-tetra-*O*-methylmannitol) of 1.94, 2.47, and 3.92 (column A, 155°), which were identified from their mass spectra¹⁵ as the acetates of 3,4,6-tri-*O*-methylmannitol, 2,3,4-tri-*O*-methylmannitol, and 2,6-di-*O*-methylgalactitol. The same components were detected on hydrolysis of the methylated polysaccharide with formic and hydrochloric acids, followed by borohydride reduction and acetylation.

The Dowex-50 column was next eluted with 15 ml of 2M ammonia in 50% aqueous methanol, the eluate was concentrated, and the residue was treated with 0.5 ml of acetic anhydride and 1 ml of pyridine at room temperature overnight. The mixture was concentrated to dryness and the residue was analysed by g l c and g l c -m s. A single peak was detected with a *T* value (relative to methyl 2-deoxy-3,4,6-tri-*O*-methyl-2-*N*-methylacetamido- α -D-galactoside) of 1.18 (column B, 210°), the mass spectrum contained the following peaks *m/e* 43 (65%), 45 (43), 56 (32), 71 (36), 73 (91), 98 (59), 100 (28), 101 (48), 102 (20), 115 (100), 142 (53), 157 (9.3), 169 (24), 186 (2.7), 187 (2.7), 214 (8), 228 (2.7), and 246 (4).

Methylation⁸ of benzyl 2-acetamido-2-deoxy-3-*O*- β -D-galactopyranosyl- α -D-glucopyranoside⁹, followed by methanolysis, isolation of the amino sugar fraction, and acetylation, afforded methyl 3-*O*-acetyl-2-deoxy-4,6-di-*O*-methyl-2-*N*-methylacetamido-D-glucopyranoside, the mass spectrum of which coincided with that of the 2-amino-2-deoxygalactose derivative isolated from the polysaccharide.

The methylated polysaccharide (~2 mg) was dissolved in 3.8 ml of methanol and 0.2 ml of 2,2-dimethoxypropane, a catalytic amount of toluene-*p*-sulphonic acid was added, and the mixture was boiled under reflux for 30 min. Sodium (~50 mg) was added and boiling was continued for 2 h. Treatment¹¹ of the reaction product with dilute acetic acid, followed by trideuteriomethylation and sugar analysis (g l c -m s of alditol acetates), revealed the formation of 1,3,5-tri-*O*-acetyl-2,6-di-*O*-methyl-4-*O*-trideuteriomethylgalactitol, *T* (relative to 1,5-di-*O*-acetyl-2,3,4,6-tetra-*O*-methylgalactitol) 1.44 (column B, 180°).

The carboxyl-reduced polysaccharide was obtained by treatment¹⁶ of a solution in methyl sulphoxide with diazomethane. The resulting methyl ester was reduced by dissolution in 0.2M borate buffer (pH 7.7) and treatment with potassium borohydride at room temperature for 1 h. The product was isolated by gel filtration.

Periodate oxidation of the polysaccharide — Polysaccharide (2.9 mg) was

treated with 3 ml of 0.1M sodium periodate for 48 h at room temperature in the dark. Potassium borohydride (100 mg) was added and, after 1 h, the reaction mixture was passed through a column (2.3 × 35 cm) of Sephadex G-25. The fraction eluted with the void volume of the column was freeze-dried and the residue was subjected to sugar analysis⁷ as follows.

The polysaccharide (~1 mg) was hydrolysed with 1 ml of 2M HCl at 100° for 4 h. The hydrolysate was concentrated to dryness, and the residue was dissolved in 0.3 ml of water and treated with 0.5 ml of 33% acetic acid and 0.5 ml of 5% aqueous sodium nitrite for 40 min at room temperature. KU-2(H⁺) resin was then added and the solution was freeze-dried. A solution of the residue in water (1 ml) was treated with 5 mg of potassium borohydride for 1 h, then deionized with KU-2(H⁺) resin, and concentrated to dryness, and boric acid was removed from the residue by distillation of methanol (3 × 2 ml) therefrom. The resulting alditols were acetylated with acetic anhydride-pyridine at 100° for 15 min, and the product mixture was subjected to g.l.c. (column A, 190°).

A solution of the oxidised-reduced polysaccharide in 2 ml of 0.5M hydrochloric acid was kept overnight at room temperature and then freeze-dried. The residue was dissolved in 2 ml of water, and treated with 30 mg of sodium metaperiodate for 16 h at room temperature and then with 20 mg of potassium borohydride for 2 h. The reaction mixture was passed in succession through columns containing 5 ml each of Amberlite IRA-401(AcO⁻) and KU-2(H⁺) resins. The eluate was concentrated to dryness and the residue was subjected to sugar analysis as described above.

Partial, acid hydrolysis of the polysaccharide — The polysaccharide (12.9 mg) was heated with 1.3 ml of 0.3M hydrochloric acid for 2 h at 100°. The solution was freeze-dried, and the residue was dissolved in water and passed through a column of Amberlite IRA-401(AcO⁻) resin (5 ml). The column was eluted with water (20 ml), and the acidic components were desorbed by elution with 10 ml of 15% acetic acid. This fraction was subjected to preparative paper electrophoresis for 90 min. Two zones (R_{GlcA} 0.7, 0.5), detected after staining of guide strips with alkaline silver nitrate, were eluted with water. Acid hydrolysis of the fast-moving component, with 2M hydrochloric acid for 16 h at 100°, afforded glucuronic acid and galactose, mannose was obtained as an additional product on hydrolysis of the second component.

A solution of the aldotriouronic acid (R_{GlcA} 0.5, ~1 μmole) in 100 μl of 0.01M borate buffer (pH 9.25) was added to a tube containing ~0.5 μmole of tritium-labelled sodium borohydride. After 2 h at room temperature, sodium borohydride (~2 mg) was added, and after a further 2 h, the solution was acidified with acetic acid and concentrated to dryness. Boric acid was removed from the residue by evaporation of methanol (5 × 2 ml) therefrom. The product was subjected to paper electrophoresis, and the acidic component (located by staining of guide strips with periodate-benzidine) was eluted with water (5 ml) to give the reduced, radioactive aldotriouronic acid. A solution of this product in 1 ml of water was treated with 70 mg of periodic acid dihydrate for 30 min, galactitol (9.66 mg, 53 μmoles) was then added followed,

after 2 h, by a solution of dimedone in phosphate buffer¹⁷ The precipitate was thrice recrystallized from methanol to give the dimedone-formaldehyde complex, m p 190–192°, activities of 5300, 4470, 4170, and 4460 counts min⁻¹ μmole⁻¹ were obtained for the crude preparation and the three crystalline samples, respectively

Determination of the configurations of the glycosidic bonds — The carboxyl-reduced polysaccharide¹⁶ (15 mg) was dissolved in freshly distilled formamide (3 ml) and treated with 2 ml of acetic anhydride and 2 ml of pyridine for 24 h at room temperature The light-brown solution was dialysed against distilled water, and the product was isolated by freeze-drying A portion of this acetylated polysaccharide was subjected to methanolysis (M hydrogen chloride in methanol, 100°, 30 min) followed by hydrolysis (2M hydrochloric acid, 100°, 4 h) The hydrolysate, after deamination, borohydride reduction, and acetylation, was analysed by g l c (column A, 190°)

The acetylated polysaccharide (~10 mg) was dried over P₂O₅ at 50°/0.1 torr for 2 h and then dissolved in 0.3 ml of glacial acetic acid, chromium trioxide (30 mg) was added, and the mixture was kept at 50° for 2 h The reaction mixture was diluted with water, and the product was extracted with chloroform The extract was washed with water and concentrated to dryness, and the residue was subjected to sugar analysis as described above

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